

**INCOMPATIBLE INSECT TECHNIQUE: A NOVEL AND
ENVIRONMENTALLY FRIENDLY TECHNOLOGY FOR
INSECT PEST POPULATION CONTROL**

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Synopsis of the Presentation

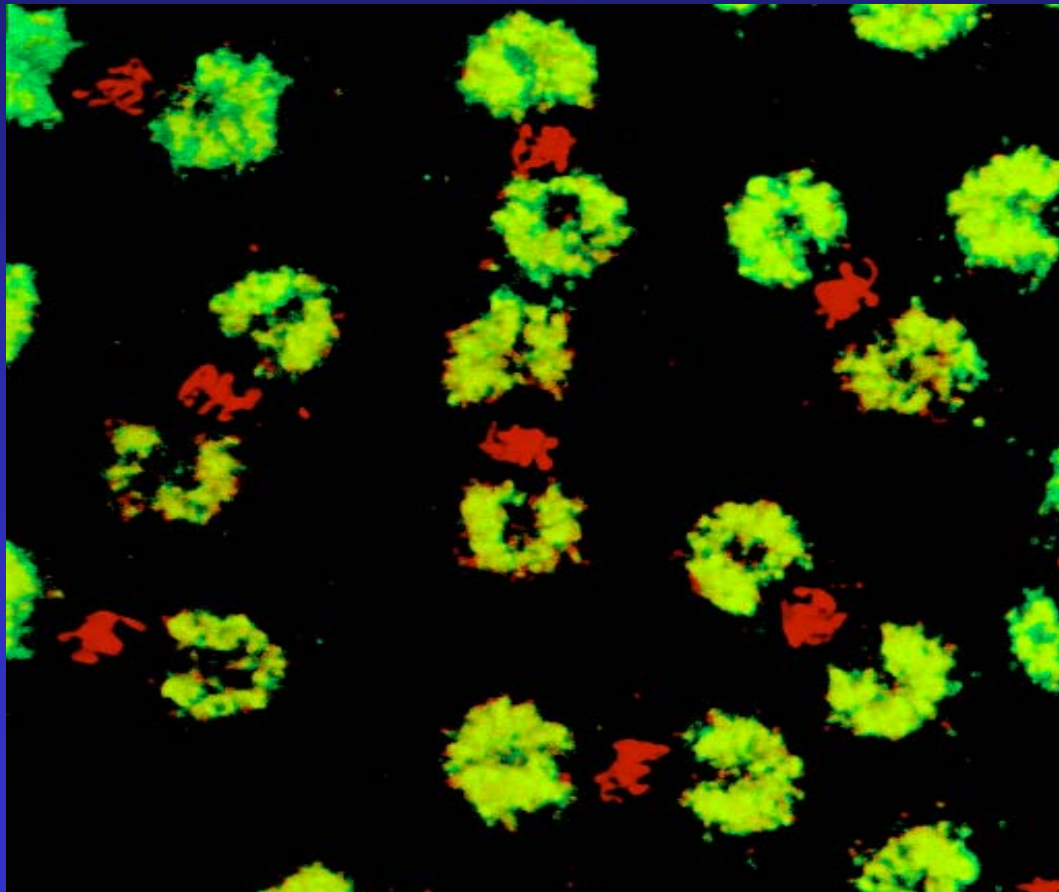
Part A: Introduction to Wolbachia Biology

Part B: Wolbachia Applications

Part C: Conclusions and Future Goals

Part A: Introduction to Wolbachia Biology

***Colbachia*: an obligatory intracellular and maternally transmitted alpha-proteobacterium that interacts with host microtubules**



Host Distribution of *Wolbachia* Strains

Arthropods

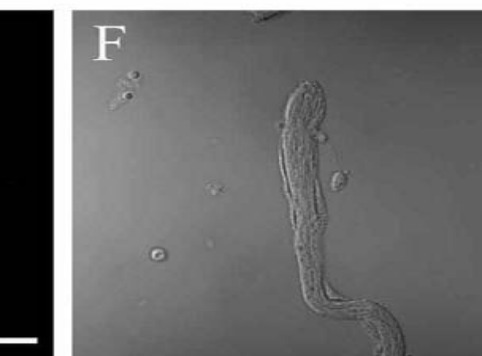
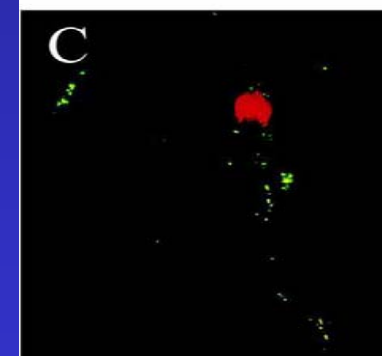
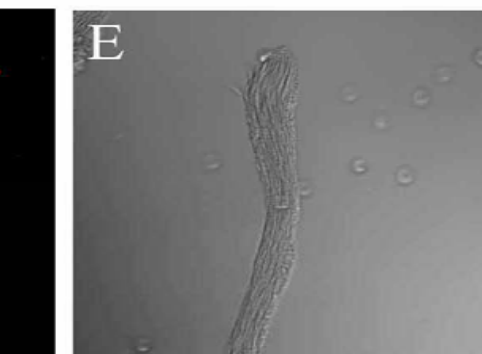
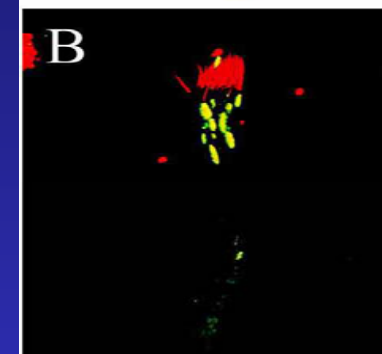
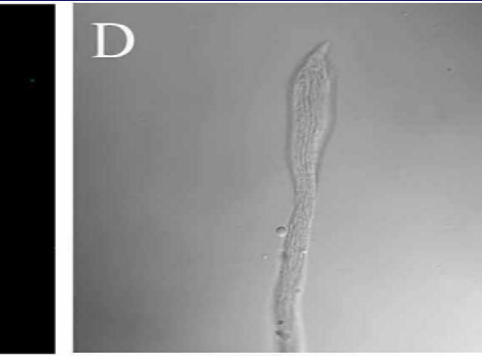
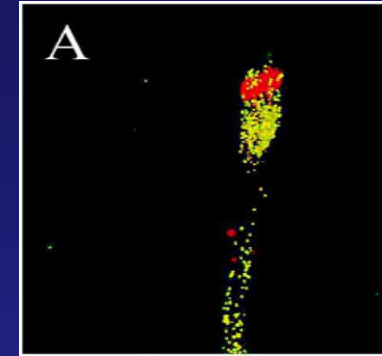
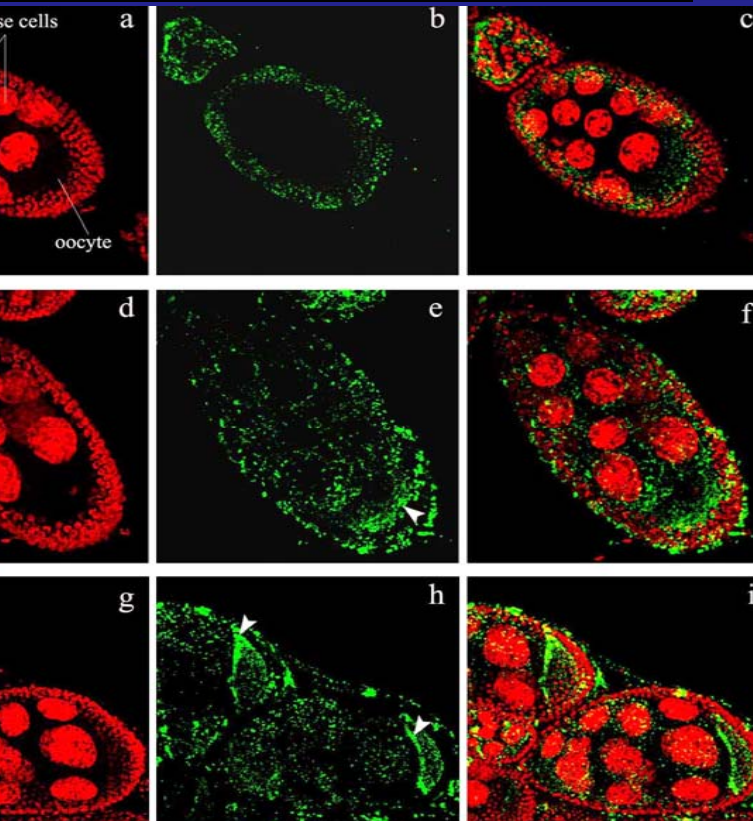
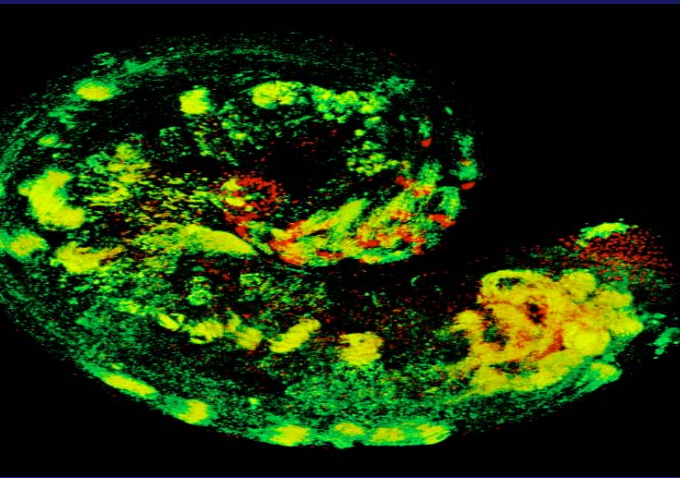
- Insects
- Mites
- Isopods
- Spiders
- Springtails

Nematodes

- Filarial worms

However, many species of agricultural importance (such as the med fly *Mediterranean fruit fly* *Ceratitis capitata* and the olive fly *Bactrocera oleae*) and species of medical importance (such as *Aedes aegypti* and *Anopheles gambiae*) are not infected with *Wolbachia*.

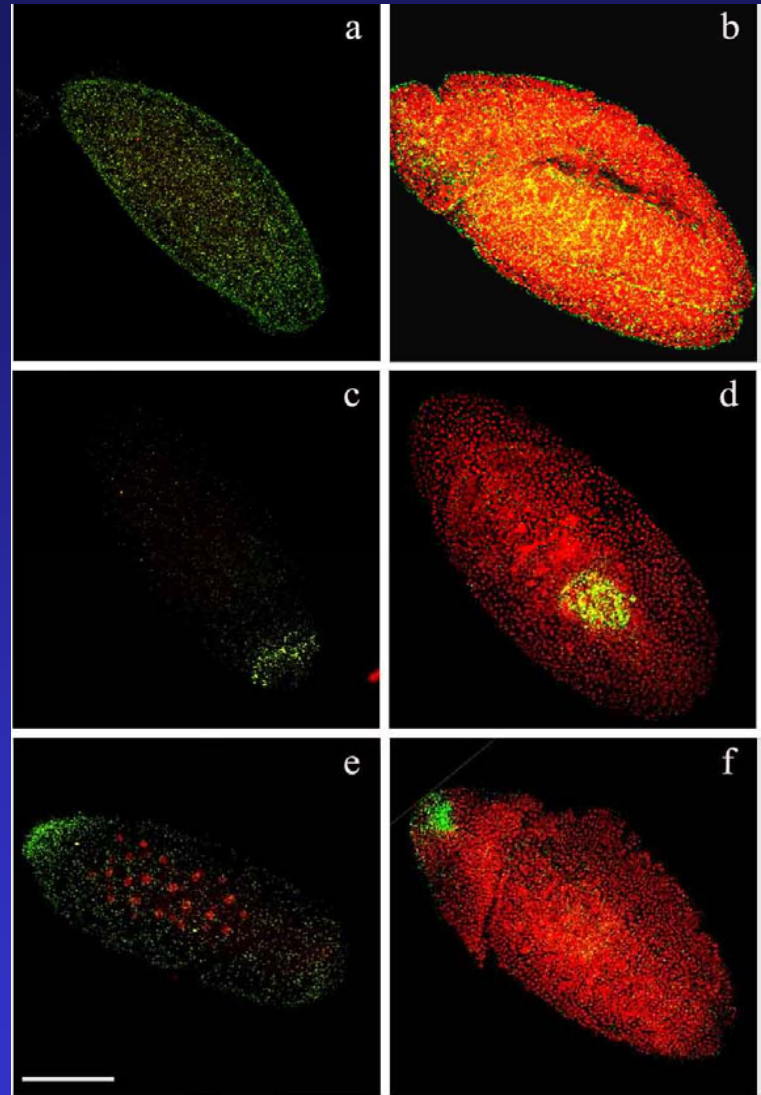
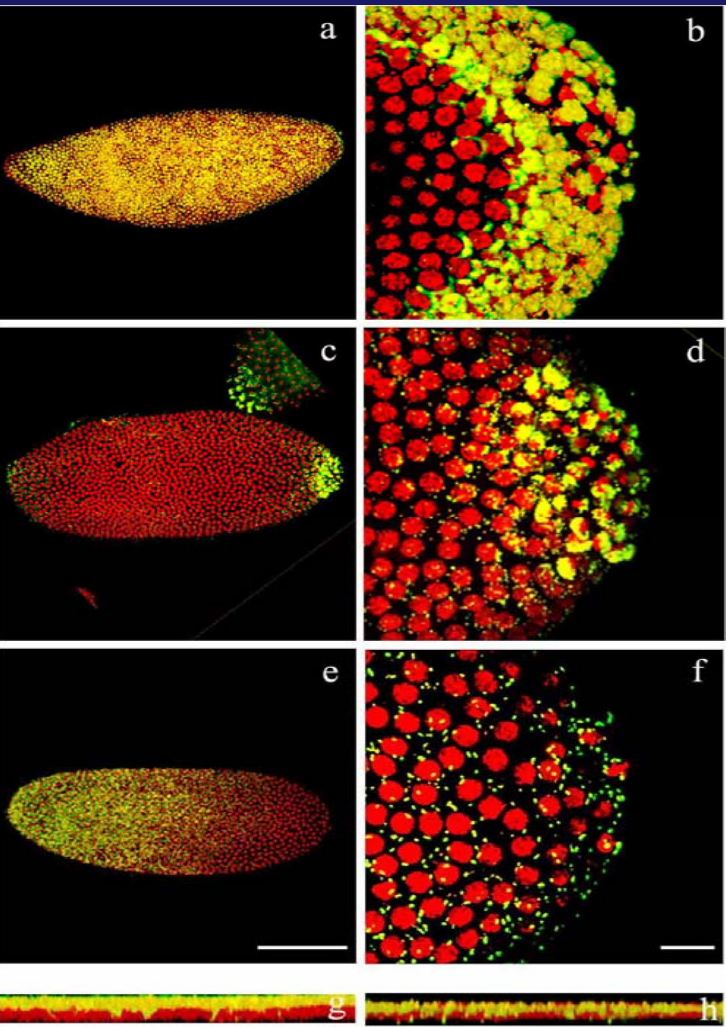
achia during *Drosophila* Spermatogenesis & Ooge



[Veneti et al., Genetics, 2003]

[Veneti et al., Appl Env Microbiol, 2003]

Wolbachia during *Drosophila* Embryogenesis



Wolbachia-Induced Reproductive Abnormalities

Wolbachia induce a number of reproductive alterations, such as:

Sexualization

Parthenogenesis

Male-killing

Cytoplasmic Incompatibility

Most of the above phenotypes “help” *Wolbachia* spreading since they favor the production of infected females.

In all cases, normal reproduction can be restored by curing the host with antibiotics, thus confirming the connection between *Wolbachia* and these phenotypes.

Uni-Directional CI

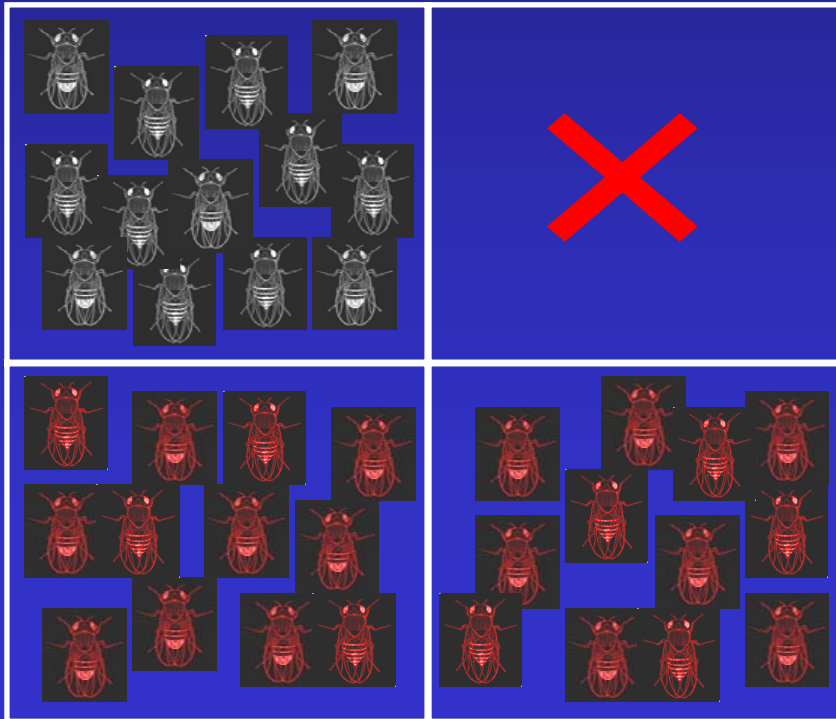
uninfected



infected



uninfected



infected



Bi-Directional CI

Infected A



Infected B



Infected A



Infected B



Major Goals of Bourtzis' Lab

- To dissect host-*Wolbachia* symbiosis towards the elucidation of the mechanism of Cytoplasmic Incompatibility (CI)
- To use the mechanism of CI for applied purposes

The EUWOL Consortium

THE EUROPEAN *WOLBACHIA* PROJECT:
TOWARDS NOVEL BIOTECHNOLOGICAL APPROACHES FOR CONTROL
OF ARTHROPOD PESTS AND MODIFICATION OF BENEFICIAL
ARTHROPOD SPECIES BY ENDOSYMBIOTIC BACTERIA

1. Bourtzis lab / Savakis lab, Greece
2. Andersson lab, Sweden
3. Garrett lab, Denmark
4. Braig lab, United Kingdom
5. Ashburner lab, United Kingdom
6. Stouthamer lab, The Netherlands [Fabrice Vavre]
7. Martin lab, France
8. Synaptic Ltd, Greece

The EUWOL Project

Genome Sequencing of:

- *w*Ri, cytoplasmic incompatibility, *Drosophila simulans*
- *w*Uni, parthenogenesis, *Muscidifurax uniraptor*
- *w*Vul, feminization, *Armadillidium vulgare*

Wolbachia Comparative Genomics

Completed Genomes: *wMel* and *wBm*

Advanced Stage: *wRi*, *wUni*, *wVul*, *wPip*, *wOvo*, *wDim*

Partial Genomes: *wSim*, *wAna*, *wWil*

SUGGESTS THAT

“core” *Wolbachia* genome is very similar between different strains (single rDNA, limited amino acid and nucleotide synthesis genes, limited number of regulatory

difference is mainly due to the different number of mobile elements, repeats and ankyrin-repeat genes

Mobile elements & ankyrin-repeat genes include the most divergent genes

Mobile genes and/or ankyrin-repeat genes may be implicated in the induction of the CI phenotype; this conclusion is also supported by the fact that the *wBm* genome contains no prophage and has a reduced number of ankyrin genes

My group is currently studying phage and ankyrin genes in a number of *Drosophila* *Wolbachia* associations exhibiting different CI phenotypes. Hopefully, we may know whether these genes are involved in the induction of CI and/or other phenotypes.

Part B: Wolbachia Applications

Wolbachia and Applied Biology

For example:

1. Asexuality
2. As an expression vector
3. As a spreading mechanism
4. As a population suppression mechanism

Wolbachia-induced Cytoplasmic Incompatibility as a tool
to suppress med fly populations



Wolbachia transfer to the med fly *Ceratitis capitata*

The entire work was done at the IMBB, Crete.

Recipient: Benakeion strain

Donor species was two populations of *Rhagoletis cerasi*:

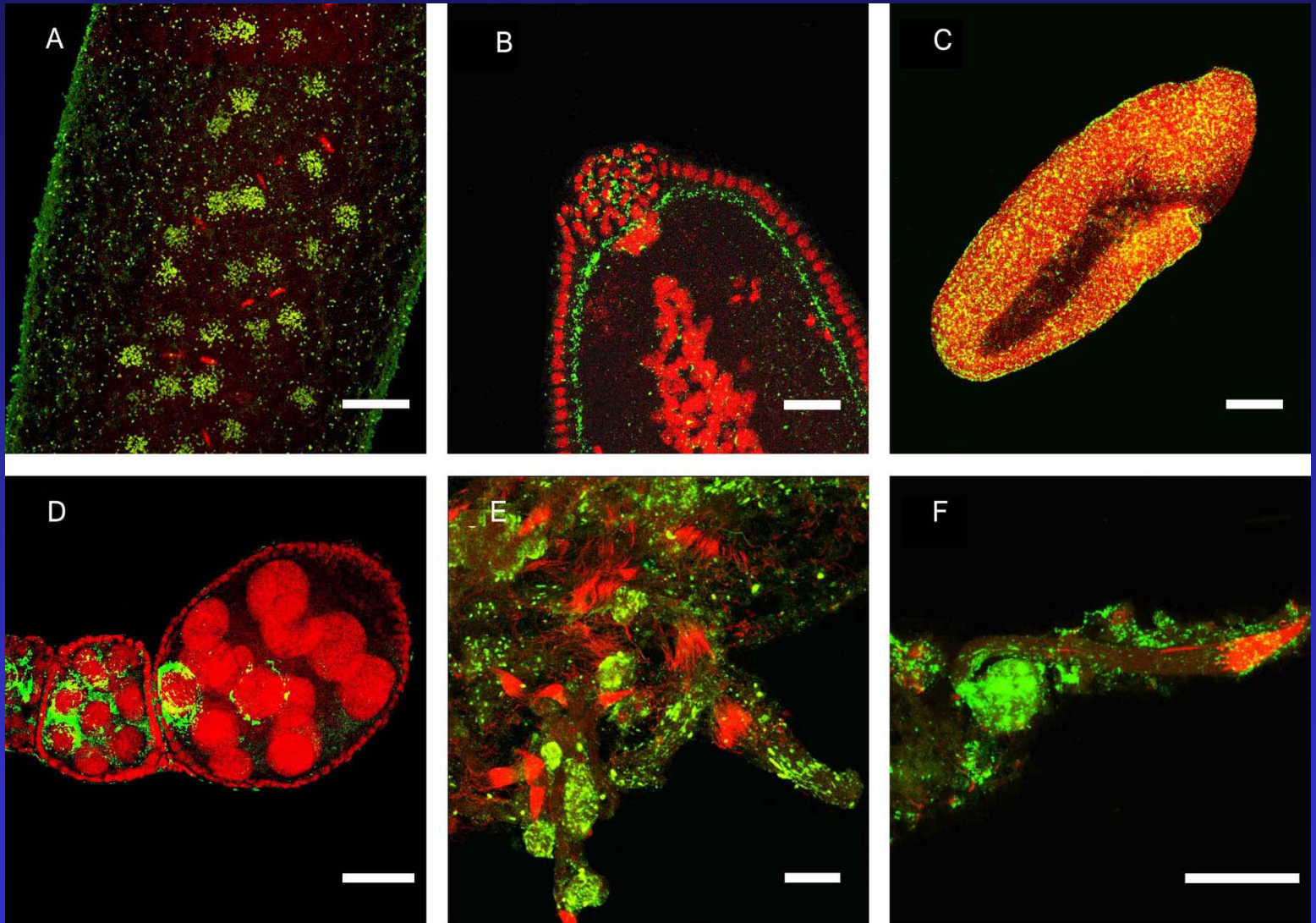
one from Austria (*wCer1* + *wCer2*) and one from Sicily (*wCer1* + *wCer4*) [supplied by M. Riegler and C. Stauffer]

Two stably transinfected lines were produced: 88.6 (*wCer2*) and S (*wCer4*).

100% infection rates (for over 3 years now, 40 generations).

[Mou et al., PNAS, 2004]

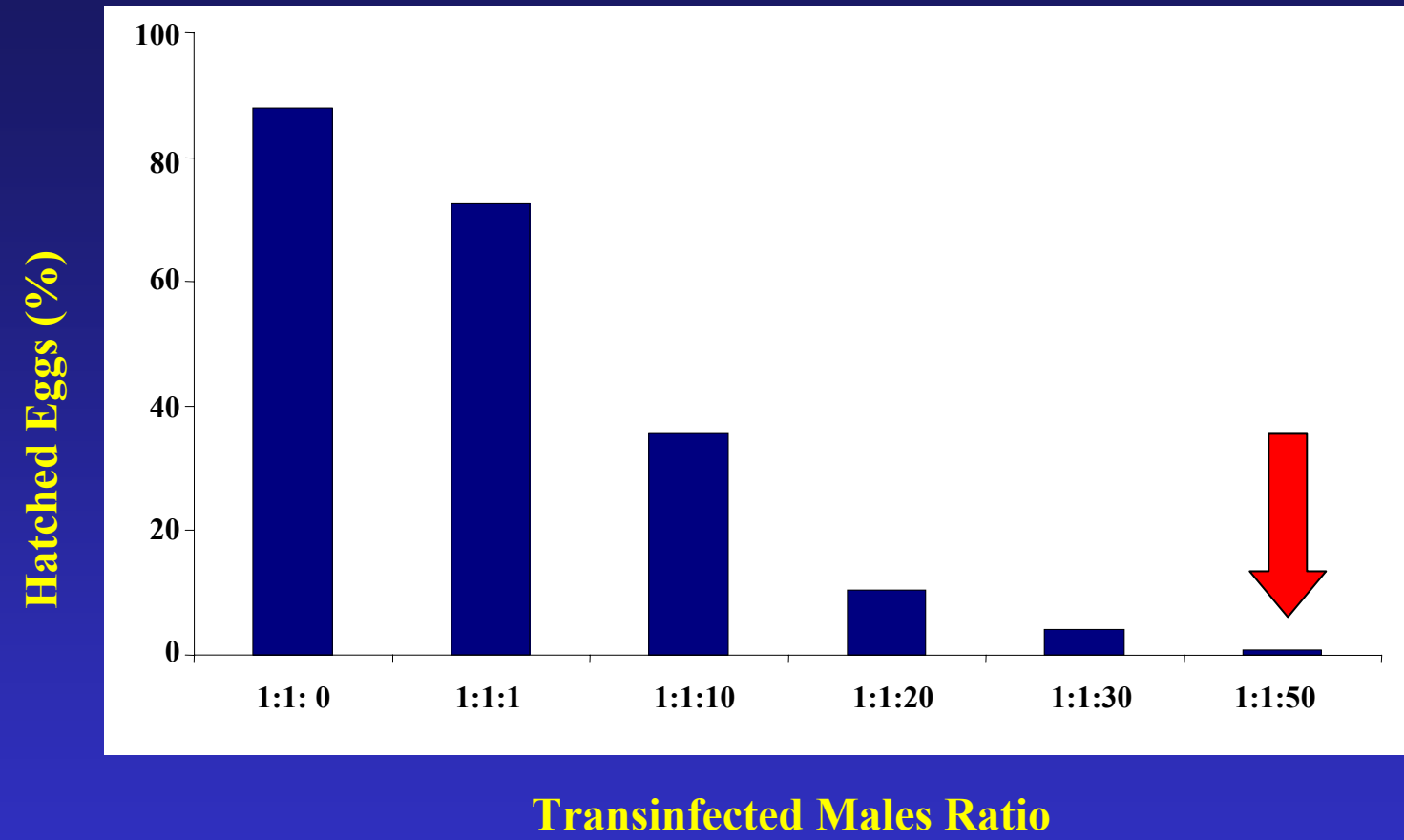
Wolbachia Distribution in Transinfected Medfly



CI assays with transinfected *Ceratitidis capitata*

<u>(females x males)</u>	<u>Embryos scored</u>	<u>Embryonic mortal</u> <u>(%)</u>
ected Benakeion x WolMed 88.6 (<i>wCer2</i>)	3000	100 ± 0
ected Benakeion x WolMed S10.3 (<i>wCer4</i>)	3000	100 ± 0
ed 88.6 (<i>wCer2</i>) x Uninfected Benakeion	3000	16.73 ± 0.68
ed S10.3 (<i>wCer4</i>) x Uninfected Benakeion	3000	32.03 ± 0.85
ed S10.3 (<i>wCer4</i>) x WolMed 88.6 (<i>wCer2</i>)	3000	100 ± 0
ed 88.6 (<i>wCer2</i>) x WolMed S10.3 (<i>wCer4</i>)	3000	100 ± 0
ed 88.6 (<i>wCer2</i>) x WolMed 88.6 (<i>wCer2</i>)	3000	64.77 ± 0.87
ed S10.3 (<i>wCer4</i>) x WolMed S10.3 (<i>wCer4</i>)	3000	67.25 ± 0.87
ected Benakeion x Uninfected Benakeion	3000	12.17 ± 0.60
WolMed S10.3 tet x WolMed S10.3 tet	1890	23.44 ± 0.97
WolMed S10.3 tet x WolMed S10.3 tet	3000	11.80 ± 0.59
WolMed 88.6 tet x WolMed 88.6 tet	2283	25.10 ± 0.91

Suppression of medfly populations using *Wolbachia*-induced



Number of adults	300	300	300	306	290	520
Number of eggs scored	3000	3000	2097	1688	858	700

Part C: Conclusions and Future Goals

Incompatible Insect Technique (I.I.T.): **Technology analogous to Sterile Insect Technique (S.I.T.)**

I.I.T. is based on the mechanism of Wolbachia-induced CI

I.I.T. is a method of genetic pest control by means analogous to S.I.T.

An effective sexing system is necessary, as is the case for S.I.T.

I.I.T. is an environmentally friendly technology

Low technological input is sufficient; Low cost technology

Higher competitiveness of released males compared to other sterility systems

I.I.T. has been used successfully in the past in field trials (for example, for the control of *Cx pipiens* in India, a project funded by WHO). At that time, it was not known that CI is induced by Wolbachia

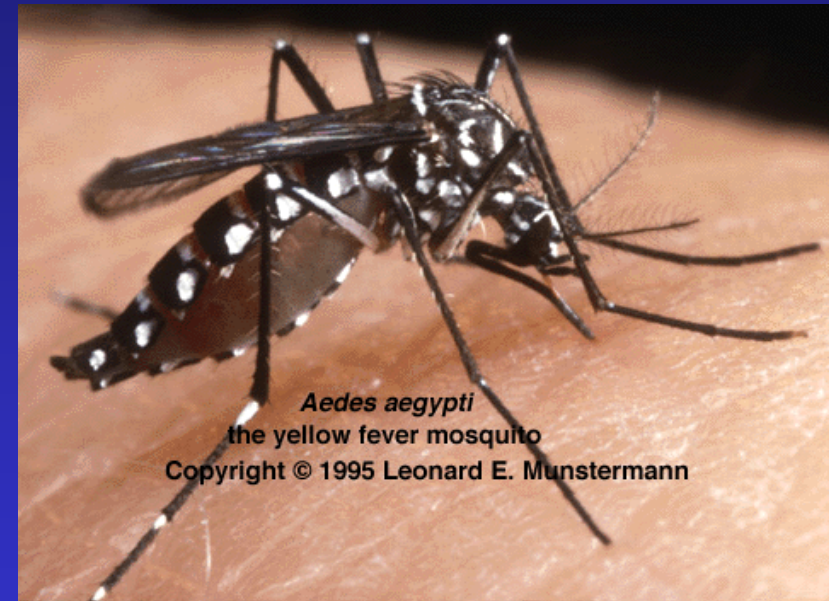
Next target species for I.I.T. applications are:

the olive fly, *Bactrocera oleae* →



Foto: Ist. Ent. Agraria

- the mosquito, *Aedes aegypti*
(then *Anopheles* sp.) ←



Aedes aegypti

the yellow fever mosquito

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We welcome suggestions for collaborations
apply the I.I.T. to your favourite insect pest species

The mechanism of CI?

Our current data,
based on comparative genomics and proteomics approaches
suggest that phage genes and/or ankyrin-repeat genes
may be implicated in the CI mechanism

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